










Anti TPO-AB

**Enzyme Immunoassay for Quantitative Determination of
Autoantibodies to Thyroid Peroxidase in Human Serum**

Instructions for use

1. SYMBOL LEGEND

| | | | |
|---|--|---|-----------------------------------|
| IVD | In vitro diagnostic medical device | LOT | Batch code |
| REF | Catalogue number |  | Manufacturer |
|  | Use by |  | Consult operating instructions |
|  | Date of manufacture |  | Biological risks |
|  | Temperature limitation | CONJ | Conjugate |
| RCNS | Reconstitute with specified volume of liquid | SUB | Substrate |
| MP | Coated microplate (96 wells) | STOP | Stop solution |
| WASH P | Wash solution, 20X concentrated | OD | Optical density |
| 20X | | DIL | Sample diluent |
| CAL | Calibrators |  | Contains sufficient for <n> tests |
| CONTROL | Control | | Deionized or distilled water |
| ASSAYB | Assay buffer | H2O | |
| TRIAL | Trial, | | |
| 5000X | 5000X concentrated | | |

2. INTENDED USE

Anti-TPO kit is provided for the **quantitative determination of autoantibodies to thyroid peroxidase (Anti-TPO) in human serum.**

TPO is a key enzyme in the biosynthesis of thyroid hormones. Anti-TPO are mainly IgG and, less frequently, IgM. In healthy individuals, concentrations of Anti-TPO in serum can reach 30 U/mL. A level exceeding this limit is the sign of an autoimmune process that usually leads to thyroid dysfunction, mainly due to reduced thyroxin secretion. Quantitative determination of Anti-TPO in serum is used for diagnostics of autoimmune thyroid diseases, such as idiopathic mixoedema, Hashimoto thyroiditis and Graves' disease.

3. PRINCIPLE OF TEST

Anti-TPO kit detects Anti-TPO of IgG class. During the first incubation, Anti-TPO from test samples and calibrators binds to TPO coated onto the inner surface of the microplate wells. During the second incubation immobilized Anti-TPO reacts with conjugate (HRP-labeled anti-human IgG). Quantity of the bound conjugate is directly proportional to Anti-TPO concentration in tested sample (Fig. 1).

During the incubation with TMB solution the colour is developing. The intensity of the colour is directly proportional to Anti-TPO concentration in specimens. Anti-TPO concentration in the patient sample is read from a standard curve that is processed in each assay.

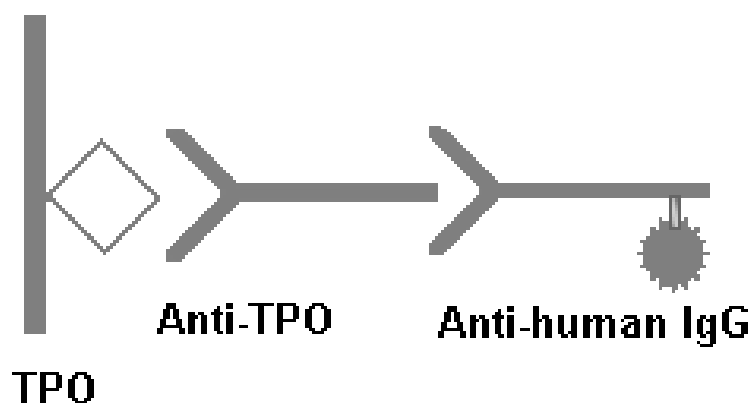


Fig. 1. Assay scheme

4. STORAGE AND STABILITY OF THE KIT

The expiration date of the kit is printed on the box label; the expiration date for each component is printed on the respective label.

Anti-TPO kit should be stored at +2...+8 °C upon receipt, preferably in the original kit box, until the expiration date. Storage at +25 °C is allowed, but for no more than 5 days.

Shelf life of the kit is 18 months.

After initial opening the kit is stable till expiry, if stored at +2...+8 °C.

If used in several separate experiments, after initial opening kit contents should be stored as follows:

- unused strips: in a firmly closed resealable zipper bag at +2...+8 °C until the expiration date;
- vials with sample diluent, conjugate, calibrators and control (ready-to-use): at +2...+8 °C for 12 months; vials with calibrators and control (reconstituted): +2...+8 °C for no more than 1 month after opening;
- vial with substrate: at +2...+8 °C until the expiration date, protected from light;
- vials with concentrated Trial, concentrated wash solution, stop solution and assay buffer: at +2...+8 °C until the expiration date;
- wash solution prepared for use: at room temperature (+18...+25 °C) for no more than 5 days or at +2...+8 °C for no more 4 weeks, in a firmly closed bottle;
- Trial solution prepared for use: at room temperature (+18...+25 °C) for no more than 5 days, in a firmly closed bottle.

Damaged Test Kits

In case of any severe damage of the test kit or components, it has to be informed in writing, during one week after receiving the kit. Usage of severely damaged single components for a test run is not recommended.

5. SAMPLE COLLECTION AND STORAGE

Collect blood by venipuncture. After clotting, the serum is separated by centrifugation.

Do not use plasma, haemolyzed (bright red) or lipaemic (milky) serum samples as well as samples containing sodium azide as preservative.

Store serum samples at +2...+8 °C for no more than 5 days. Aliquot and freeze samples for longer storage (−20 °C and lower). Avoid repeated freezing.

6. EXPECTED VALUES

324 serum samples collected between 9–11 a.m. from healthy individuals at the age 21-45 were assayed with Anti-TPO kit. In 99.4 % of patients Anti-TPO concentration did not exceed 30 U/mL. This limit should be considered as guidelines only.

It is highly recommended for each laboratory to determine its own reference range of Anti-TPO concentrations.

7. QUALITY CONTROL

It is recommended to use control samples according to the state and federal regulations. The use of control samples is advised to assure the day to day validity of results.

8. REAGENT PREPARATION

Allow all the reagents to reach room temperature (+18...+25 °C), and then thoroughly stir.

MP Keep **microplate** at room temperature for at least 30 minutes before opening the bag. Place required number of strips onto strip holder. Place unused strips into the resealable zipper bag and reseal duly.

CAL **CONTROL** Calibrators and Control

Liquid calibrators and control are ready to use.

Prepare lyophilized calibrators and control as follows. Gently tap on the vial caps to knock off all the dry matter. Open the vials and carefully place the caps upside down on the clean dry surface. Add 0.5 mL of

distilled or deionized water to each vial with lyophilized calibrators and control, close vials with the corresponding caps and leave for 10 min at room temperature without stirring. Then stir gently avoiding foaming, until the dry matter is completely dissolved. Leave for another 10 minutes at room temperature stirring gently periodically. Make sure that no dry matter is left on the caps and walls of the vials.

WASH P Prepare required volume of **wash solution** by dilution of the concentrate 20-fold with distilled or deionized water. For example:

5 mL of **WASH P 20X** + 95 mL of water.

Mix thoroughly, avoid foaming.

SUB Protect **substrate** from direct light.

9. SAMPLE PREPARATION

Allow samples to reach room temperature (+18...+25 °C). Stir samples gently in order to ensure homogeneity.

Before the analysis all the serum samples (except calibrators and control) should be diluted 100-fold with sample diluent **DIL** in concordance with instructions for use for manually before analysis. The example of manual sample dilution as follows (Sample 1). If expected Anti-TPO concentration in the sample 1 is higher than in **calibrator 5**, it should be diluted 20-fold with sample diluent **DIL** (Sample 2).

Sample 1 (100-fold dilution): **990 µL** of sample diluent **DIL** and **10 µL** of serum sample.

Sample 2 (additional 20-fold dilution): **190 µL** of sample diluent **DIL** and **10 µL** of **sample 1**.

10. PERFORMANCE CHARACTERISTICS OF THE ASSAY

10.1. Calibration-Traceability

Anti-TPO test kit was calibrated against internationally approved NIBSC Research Standard 66/387.

10.2. Specificity

High specificity of the assay is ensured by high purity TPO that was used for manufacture of the kit.

10.3. Analytical Sensitivity

Analytical sensitivity of Anti-TPO assay, i.e. concentration that can be distinguished from calibrator 0, is 4 U/mL. It is defined as mean OD of 10 replicates of calibrator 0 plus two SD standard deviations.

10.4. Measurement Range

Anti-TPO kit was validated for measurement of Anti-TPO concentration within the concentration diapason (without dilution) of 4-500 U/mL”).

10.5. Intra- and Inter-Assay Variation

10.5.1. Precision (intra- and inter-assay CV) for manual kit

Intra-assay CV

To determine intra-assay CV 8 serum samples were assayed in 9 replicates each. The results are shown below.

| Sample | Mean concentration of Anti-TPO, U/mL | Intra-assay CV | |
|--------|--------------------------------------|----------------|-------|
| | | SD | CV, % |
| HS 1 | 22.945 | 0.4 | 1.6 |
| HS 2 | 51.285 | 3.6 | 7.0 |
| HS 3 | 100.91 | 0.8 | 0.9 |
| HS 4 | 234.27 | 12.7 | 5.4 |
| HS 5 | 383.27 | 9.1 | 2.4 |
| HS 6 | 78.033 | 2.6 | 3.3 |
| HS 7 | 34.335 | 2.1 | 6.1 |
| HS 8 | 444.85 | 28.2 | 6.3 |

Inter-assay CV

To determine inter-assay CV 8 serum samples were assayed 3 times by different operators with 1-week interval. Each specimen was assayed in 9 replicates. The results are shown below.

| Sample | Mean concentration of Anti-TPO, U/mL | | | Inter-assay precision | |
|--------|--------------------------------------|---------|---------|-----------------------|-------|
| | 1 assay | 2 assay | 3 assay | SD | CV, % |
| HS 1 | 34.3 | 33.8 | 34.3 | 0.3 | 0.9 |
| HS 2 | 23.1 | 23.5 | 26.0 | 1.6 | 6.5 |

| | | | | | |
|------|-------|-------|-------|-----|-----|
| HS 3 | 96.0 | 93.4 | 103.3 | 5.1 | 5.2 |
| HS 4 | 134.3 | 123.3 | 133.3 | 6.0 | 4.6 |
| HS 5 | 155.5 | 146.8 | 156.1 | 5.2 | 3.4 |
| HS 6 | 157.8 | 146.8 | 147.1 | 6.3 | 4.2 |
| HS 7 | 101.2 | 92.0 | 102.1 | 5.6 | 5.7 |
| HS 8 | 318.0 | 315.7 | 307.0 | 5.8 | 1.8 |

11. LIMITATION OF THE METHOD

Any clinical diagnosis should not be based on the results of in vitro diagnostic methods alone. For diagnosis establishment, a physician is supposed to consider all available clinical and laboratory findings.

12. SAFETY PRECAUTIONS

- **This kit is for in vitro diagnostic use only.** Operator should follow the manual closely in order to ensure reliable data. The manual is valid for the present kit only, within the listed composition. Any substitution of kit components is not allowed by CE regulations.

- Do not use kit or its components after the expiration date indicated on the label. Take into account stability period for reconstituted reagents.

- Do not mix or use together reagents from different lots of the kit except substrate, stop solution and wash solution.

- Do not use substrate, stop solution and wash solution supplied by other vendors.

- Use only "P"-labeled wash solution.

- Note that stop solution is 1 N HCl solution. Avoid contacts with skin and mucosa. In case of contact rinse affected area thoroughly with plenty of water and seek medical advice.

- Take into account the following common procedural notes:


- always pipette reagents into wells immediately after washing procedure;


- avoid contamination of the solutions;


- in case of partial use of the kit, dispense only required volume of the reagent into the tray;


- do not pour unused reagents back into the original vials;
- avoid exposure to direct sunlight during incubations;
- always pipette reagents in the same order to minimize reaction time differences between wells; the total dispensing time for the calibrators, control and samples must not exceed 15 min;
- the incubation temperature for all the immunological reactions must be kept at +37 °C;
- do not touch the bottom of the wells;
- calibrators should be measured in each separate assay. It is also recommended to measure each time Anti-TPO concentration in the control.

- TMB solution should be colourless. Light colouring of solution is admissible. Avoid direct exposure of substrate to sunlight.

-  Source materials of human origin used for kit components preparation were tested and found negative for HBsAg, anti-HIV and anti-HCV antibodies. However, none of known laboratory test guarantees absence of these viral agents. Therefore, all kit components and patient's samples should be handled as potentially hazardous.

-  After usage strips, calibrators, control, specimens and all consumables which contacted with specimens during handling, storage or assay (tubes, vials, gloves, pipette tips etc.) should be collected separately and sterilized by autoclaving. Instead of autoclaving pipette tips may be sterilized by disinfectant treatment. After sterilization all components and expendable materials may be utilized as non-dangerous garbage. Other components of the kit should be discarded into conventional garbage.

-  During manual washing procedure do not discard the contents of the wells directly to drainage. Use a container with disinfectant solution.

-  As the kit contains potentially hazardous material, the following precautions should be taken:

- do not smoke, eat or drink while performing the assay;
 - always use protective gloves;
 - never pipette material by mouth;
 - in case of spilling, wipe up the spills promptly and wash affected area thoroughly using decontaminant.
- GLP including all general and individual regulations should be applied for the kit usage.



As the kit contains irritant (**CONJ**, **CAL**, **DIL**, **CONTROL**), the following precautions should be observed:

- P261 - Avoid breathing spray;
- P272 - Contaminated work clothing should not be allowed out of the workplace;
- P280 - Wear protective gloves/protective clothing/eye protection;
- P302+P352 - IF ON SKIN: Wash with plenty of soap and water;
- P333+P313 - If skin irritation or rash occurs: Get medical advice/attention;
- P363 - Wash contaminated clothing before reuse;
- P501 - Dispose of contents/container in accordance with national regulation.

Precautionary statements according to Regulation EC № 1272/2008.

13. MATERIAL PROVIDED

13.1. Material Provided

| | | |
|-----------------------|---|--|
| MP | Microplate: 12 breakable 8-well strips (total 96 wells) coated with human TPO | 1 pcs |
| CONJ | Conjugate: solution containing monoclonal antibodies against human IgG conjugated with HRP | 16 mL, ready to use |
| 0-5 CAL | Anti-TPO calibrators: protein-based solution containing known Anti-TPO concentrations – 0; 25; 50; 100; 250; 500 U/mL (approximate values). The concentrations of calibrators may be different for schemes with or without shaking. For exact Anti-TPO concentrations, see vial labels | 6 vials , 0.5 mL each; ready to use or lyophilized preparations |
| CONTROL | Anti-TPO control: protein-based solution or lyophilized preparation containing known Anti-TPO concentration. The range of Anti-TPO concentration may be different for schemes with or without shaking. For exact range of Anti-TPO concentration, see vial label | 0.5 mL, ready to use or lyophilized preparation |
| SUB | Substrate (TMB solution): 3,3',5,5'-tetramethylbenzidine solution in citrate buffer containing hydrogen peroxide | 14 mL, ready to use |
| WASH P 20X | Wash solution P, 20X concentrated: surfactant in buffered saline, sufficient for preparation of 840 mL of solution | 3 x 14 mL, concentrated |
| STOP | Stop reagent: 1 N HCl solution | 14 mL, ready to use |
| ASSAYB | Assay buffer | 14 mL, ready to use |
| DIL | Sample diluent | 50 mL, ready to use |

13.2. Equipment and Materials Required but not provided

- 1-channel calibrated variable precision pipettes, with disposable tips;
- 8-channel calibrated variable precision pipette, with disposable tips;
- microplate incubatorb (+37 °C) or microplate incubator/shaker (+37 °C, shaking speed 500–800 rpm);
- manual or automatic equipment for rinsing wells;
- calibrated microplate reader (450 nm);
- vortex tube mixer;
- deionized or distilled water;
- graduated beaker and cylinder of appropriate volume;
- latex or plastic gloves;
- trays for pipetting reagents with 8-channel pipette;
- disinfectant;
- absorbent material (for manual wash).

13.3. Test Procedure

Anti-TPO kit is designed for 96 tests. This is sufficient for the **quantitative assay** of 40 unknowns, 6 calibrators, 1 control and 1 blank (OD of TMB solution) in duplicates, provided that all the strips are used simultaneously.

13.3.1. Assay Procedure

13.3.1.1. Protocol with shaking

(See assay scheme, section 13.5)

A. Pipette

- 100 µL of assay buffer **ASSAYB** into each well,

Leave wells A1-A2 empty for blank!

B. Pipette:

- 50 µL of calibrators **CAL** (0-5), control **CONTROL** and diluted patient's samples in duplicates into the respective wells;

Note: total time of dispensing must not exceed 15 minutes, otherwise the test result may be unreliable, because the time of incubation will substantially vary for different samples.

C. Incubate strips for 30 minutes while shaking (500–800 rpm) at +37 °C.

D. Wash 5 times, as described in section 13.3.2.

E. Pipette:

- **120 µL of conjugate [CONJ]** into each well, **except wells A1-A2.**

F. Incubate strips for 30 minutes while shaking (500–800 rpm) at +37 °C.

G. Wash 5 times, as described in section 13.3.2.

H. Pipette 100 µL of substrate [SUB] into each well (including blank); incubate **at room temperature (+18...+25 °C) in the dark for 15-30 minutes**, depending on the colour intensity, or **10 minutes while shaking (500-800 rpm) at +37 °C.**

I. Pipette 100 µL of stop solution [STOP] into each well (including blank) in the same sequence and at the same speed as used for dispensing TMB substrate. Shake for 1–2 min at room temperature.

J. Read OD at 450 nm within 20 min.

13.3.1.2. Protocol without shaking

(See assay scheme, section 13.6)

A. Pipette

- **100 µL of assay buffer [ASSAYB]** into each well,
Leave wells A1-A2 empty for blank!

B. Pipette:

- **50 µL of calibrators [CAL]** (0-5), **control [CONTROL]** and diluted patient's samples in duplicates into the respective wells;
Note: total time of dispensing must not exceed 15 minutes, otherwise the test result may be unreliable, because the time of incubation will substantially vary for different samples.

C. Incubate strips for 45 minutes at +37°C (pre-shake for 1-2 minutes at room temperature).

D. Wash 5 times, as described in section 13.3.2.

E. Pipette:

- **120 µL of conjugate CONJ into each well, except wells A1-A2.**

F. Incubate strips for 45 minutes at +37°C.

G. Wash 5 times, as described in section 13.3.2.

H. Pipette 100 µL of substrate SUB into each well (including blank); incubate at room temperature (+18...+25°C) in the dark for 15-30 minutes, depending on the colour intensity.

I. Pipette 100 µL of stop solution STOP into each well (including blank) in the same sequence and at the same speed as used for dispensing TMB substrate. Shake for 1–2 min at room temperature.

J. Read OD at 450 nm within 20 min.

13.3.2. Wash Procedure

It is advisable to use an automatic microplate washer set at 5 wash cycles and a volume of 300 µL of wash solution per well per cycle.

If an automatic washer is not available, the wash procedure can be carried out manually as follows :

- remove the contents of the wells into a container with disinfectant;
- dispense 300 µL of wash solution (prepared according to section 8) into each well, shake the plate carefully for 5–10 sec and remove the contents of the wells; repeat 5 times;
- strike the wells sharply on absorbent material to remove any liquid residue.

13.4. Data Processing

If the reader cannot be adjusted to zero using the substrate blank in wells A1-A2, subtract mean OD value of wells A1-A2 from all OD values before further calculations.

Example:

OD (Cal 5) measured = 2.28 and OD (blank) = 0.06;

OD (Cal 5) calculated = $2.28 - 0.06 = 2.22$

13.4.1. Data Reliability (for OD Measured at 450 nm)

The data should meet the following criteria:

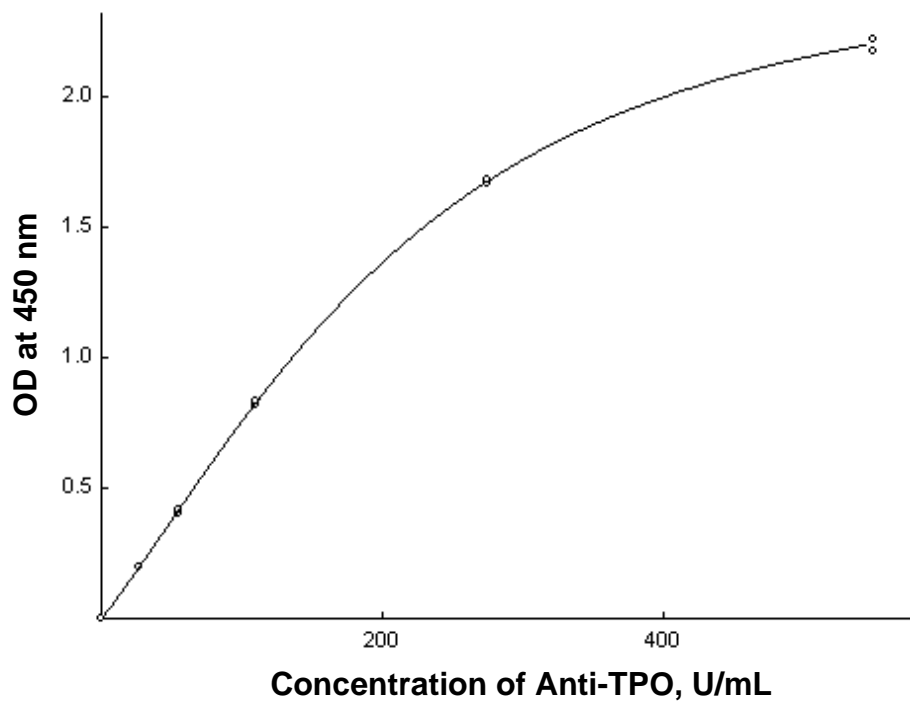
- average blank OD (in wells A1-A2) ≤ 0.100 ;
- average OD of Cal 5 ≥ 1.0 (after blank subtraction);
- control's concentration must fall within the acceptability range that is shown on the vial label.

If the data obtained do not meet the criteria, the results are considered unreliable and the test should be repeated.

13.4.2. Quantitative Determination

Specialized software for quantitative determination is recommended. Mean OD of calibrators are plotted versus their respective Anti-TPO concentrations using 4PL or 5PL fit (see typical standard curve, fig. 2). Calculate concentration of Anti-TPO in samples using standard curve.

Any extrapolation of the standard curve to Anti-TPO concentration above the nominal value of **calibrator 5** is forbidden. In this case, the sample should be additionally diluted 20-fold with sample diluent and re-tested. Multiply the measured concentration of additionally pre-diluted samples by additional dilution factor (20-fold).



**Fig. 2. Example of typical standard curve.
Do not use for evaluation of real assay data!**

13.5. Assay scheme with shaking

| Reagents | Wells | «Blank» | CAL CONTROL | Samples |
|------------------------------|--------------------------------------|---------|------------------------------|---------|
| | ASSAYB | – | – | 100 µL |
| CAL CONTROL | – | – | 50 µL | – |
| Samples | – | – | – | 50 µL |
| Incubation No. 1 | 30 min, +37 °C, 500–800 rpm | | | |
| WASH P (diluted) | 5 x 300 µL | | | |
| CONJ | – | – | 120 µL | 120 µL |
| Incubation No. 2 | 30 min, +37 °C, 500–800 rpm | | | |
| WASH P (diluted) | 5 x 300 µL | | | |
| SUB | 100 µL | – | 100 µL | 100 µL |
| Incubation No. 3 | 15–30 min, +18...+25 °C, in the dark | | | |
| | 10 min, +37 °C, 500–800 rpm | | | |
| STOP | 100 µL | – | 100 µL | 100 µL |
| Stirring | 1–2 min, +18...+25 °C | | | |
| OD measuring | 450 nm | | | |
| Calculations | Corresponding software | | | |

13.6. Assay scheme without shaking

| Reagents | Wells | «Blank» | CAL | Samples |
|-------------------------|---|---------|----------------|---------|
| | | | CONTROL | |
| ASSAYB | – | – | 100 µL | 100 µL |
| CAL | – | – | 50 µL | – |
| CONTROL | – | – | – | – |
| Samples | – | – | – | 50 µL |
| Incubation No. 1 | 45 min, +37°C (pre-shake for 1-2 minutes at room temperature) | | | |
| WASH P (diluted) | 5 x 300 µL | | | |
| CONJ | – | – | 120 µL | 120 µL |
| Incubation No. 2 | 45 min, +37°C | | | |
| WASH P (diluted) | 5 x 300 µL | | | |
| SUB | 100 µL | 100 µL | 100 µL | 100 µL |
| Incubation No. 3 | 15–30 min, +18...+25°C, in the dark | | | |
| STOP | 100 µL | 100 µL | 100 µL | 100 µL |
| Stirring | 1–2 min, +18...+25°C | | | |
| OD measuring | 450 nm | | | |
| Calculations | Corresponding software | | | |

BIOGENIX INC. PVT. LTD.

Factory: B - 19/A, S.I.L Ancillary Estate, Amausi Industrial Area, Nadarganj, Kanpur Road, Lucknow - 226008 (U.P.), India

Email: biogenix2007@yahoo.com, info@biogenixinc.com

Website: www.biogenixinc.com

Customer care no: +919140971443